

Wahlpflichtfach „Vertiefung Physikalische Chemie“  
Masterstudiengang Modul WP04

Moderne Methoden der Spektroskopie

## **Prinzip Microarrays und Biosensoren**



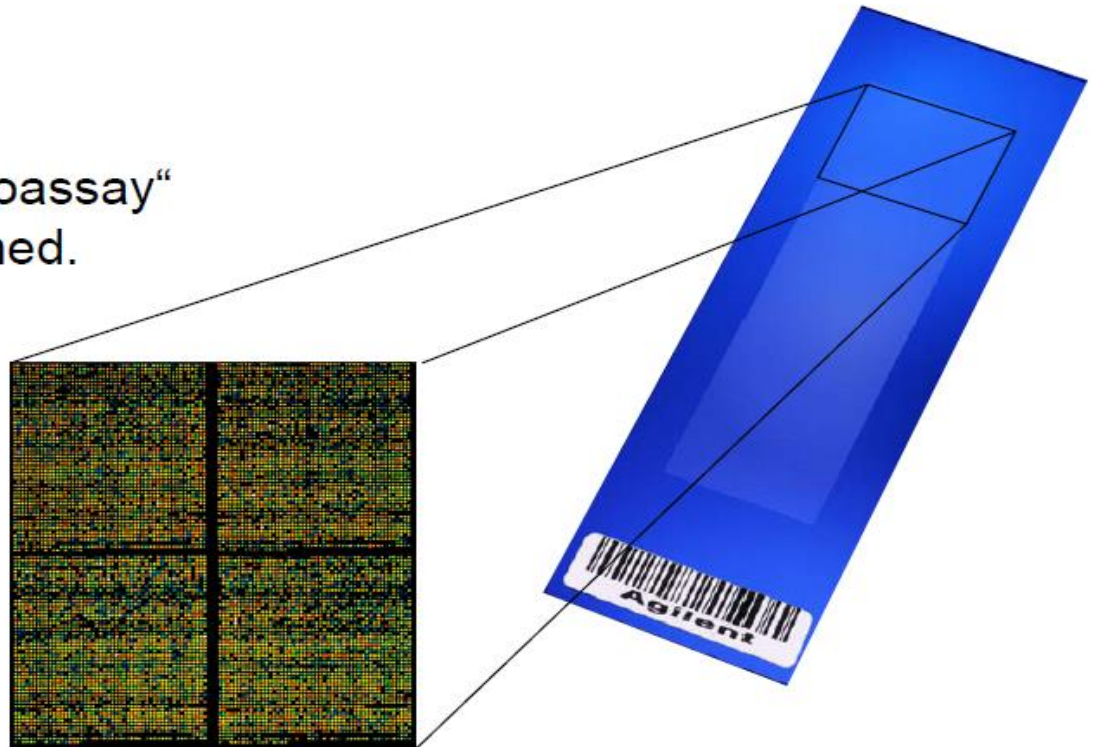
BEUTH HOCHSCHULE FÜR TECHNIK BERLIN  
University of Applied Sciences

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# What is a Microarray?

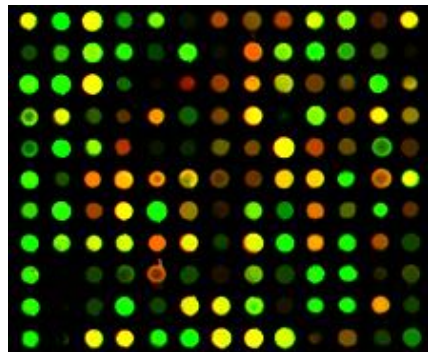
A microarray is a spatially ordered, minituarized arrangement of a multitude of immobilized reagents

First publication:  
Roger Ekins et. al.,  
„Multi-analyte immunoassay“  
1989, J. Pharm. Biomed.  
Anal. 7: 155 - 168

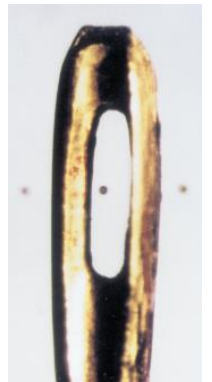
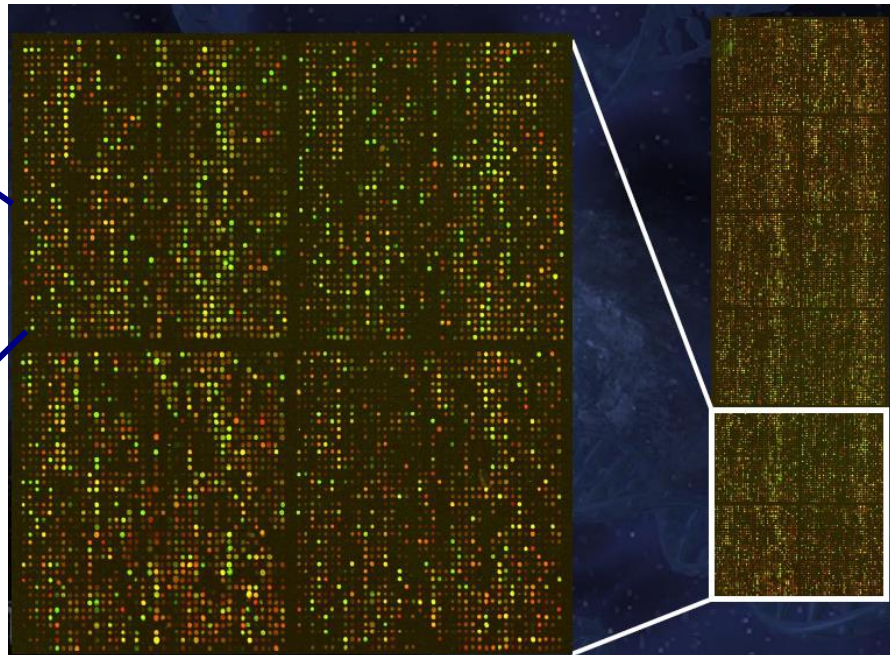


# Why DNA Microarrays?

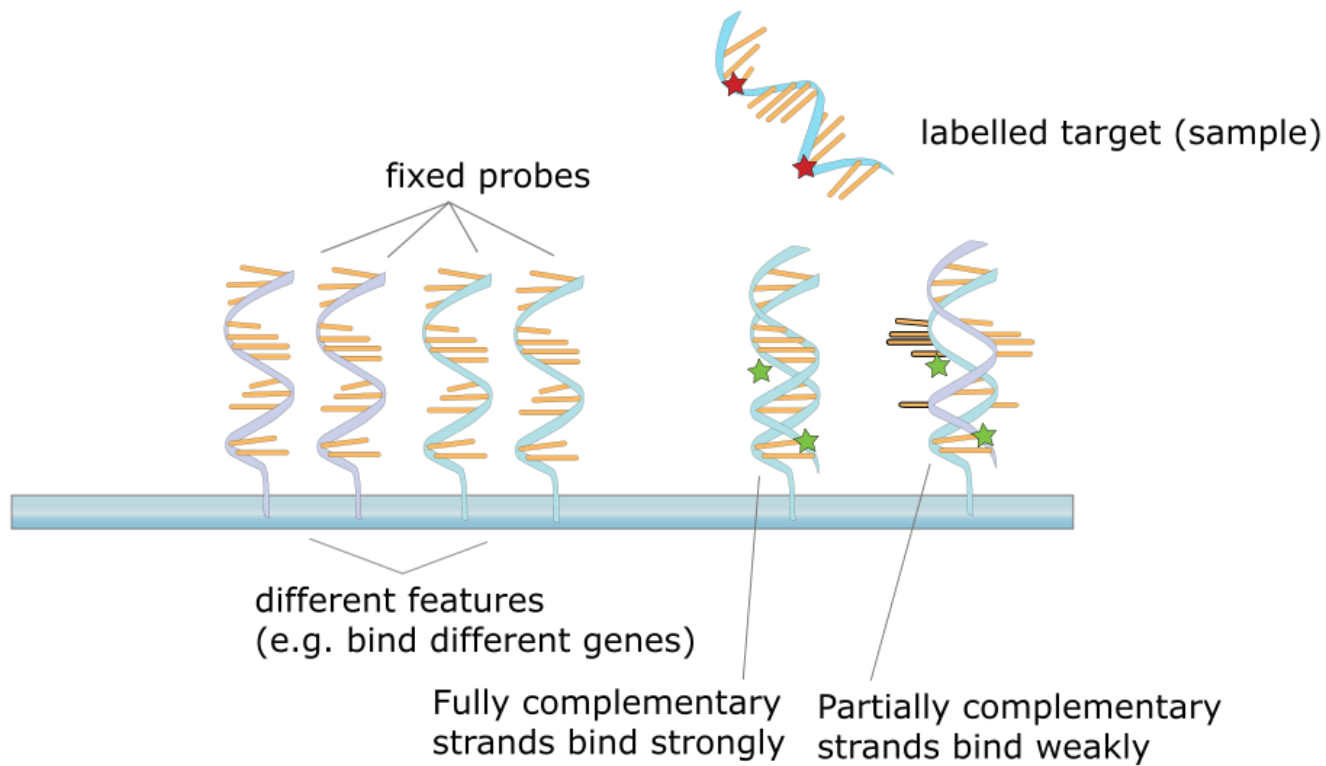
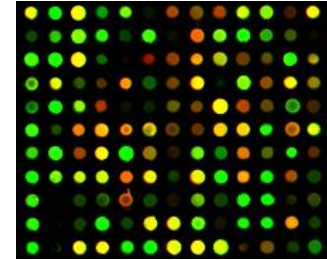
- Genomics → 40-80000 Genes
- Gene expression (Mutation, Polymorphism) of single cell populations
- Miniaturization → simultaneous analysis  
→ High Throughput Analysis ⇒ Microarrays
- Nylon Membranes 80-100 spots/cm<sup>2</sup>
- DNA Chips up to 10000 spots/cm<sup>2</sup>
  - Size of a Spot > 500 μm → Macroarray, < 500 μm → Microarrays



Spot: 50 – 200 μm

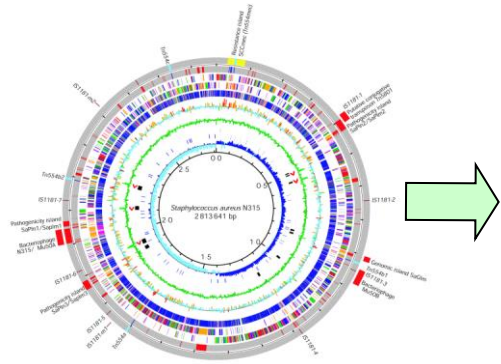


# The Principle of DNA Microarrays

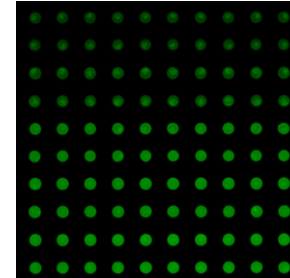
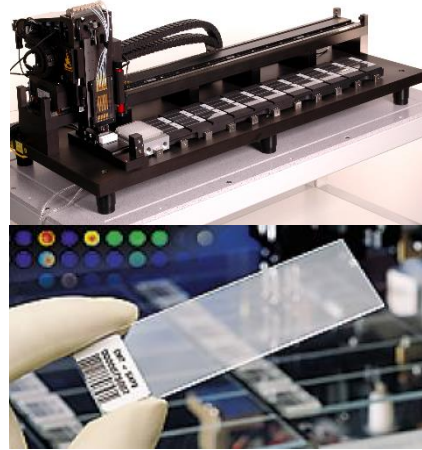


# DNA-Microarrays → Workflow

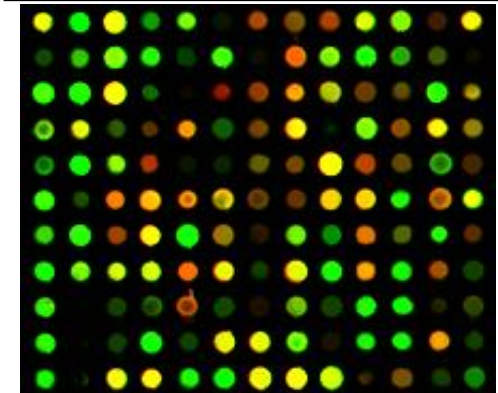
Bioinformatic → Probe Design+ Synthesis



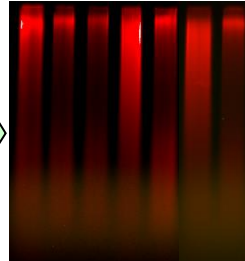
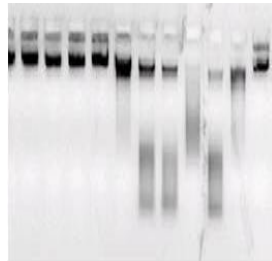
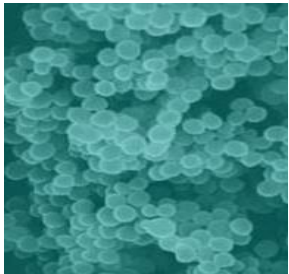
Array Printing+ Immobilization



Hybridization, Scannen



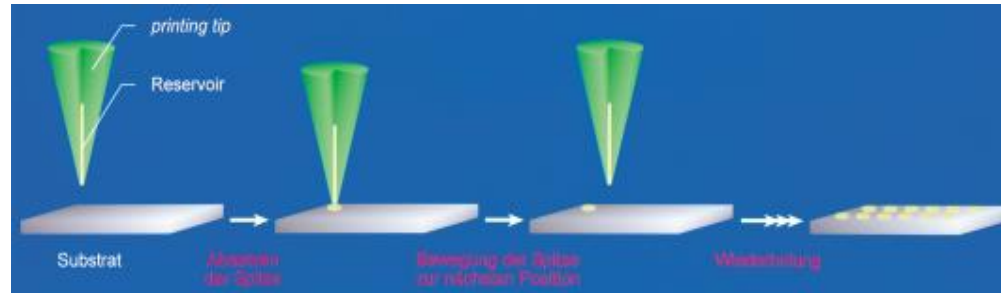
Sample preparation → Extraction of Nucleic Acids, Amplification/ Labeling of DNA/ RNA



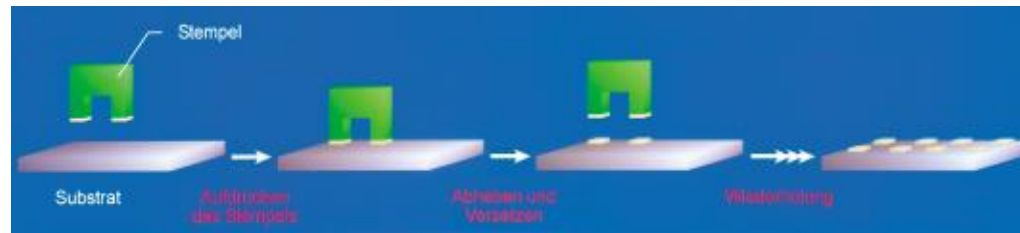
Data Analysis

# Deposition Techniques (Printing, Spotting)

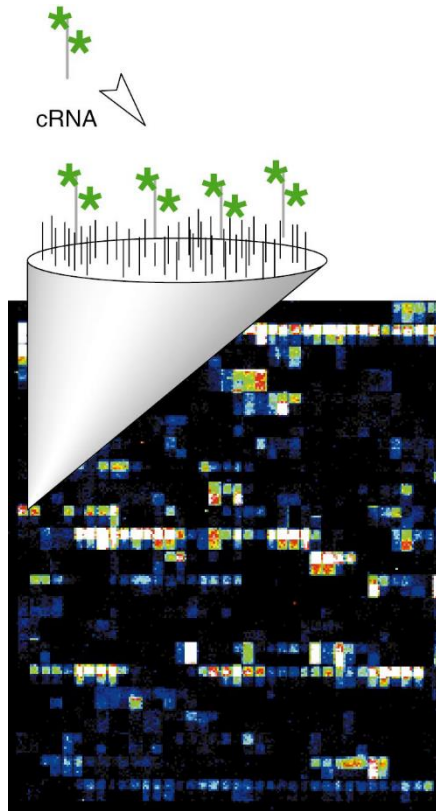
## Contact tip



## Microcontact printing



# Microarrays for Gene Expression Profiling



Hybridization of immobilized probe DNA of known sequence with target DNA carrying a fluorescent marker

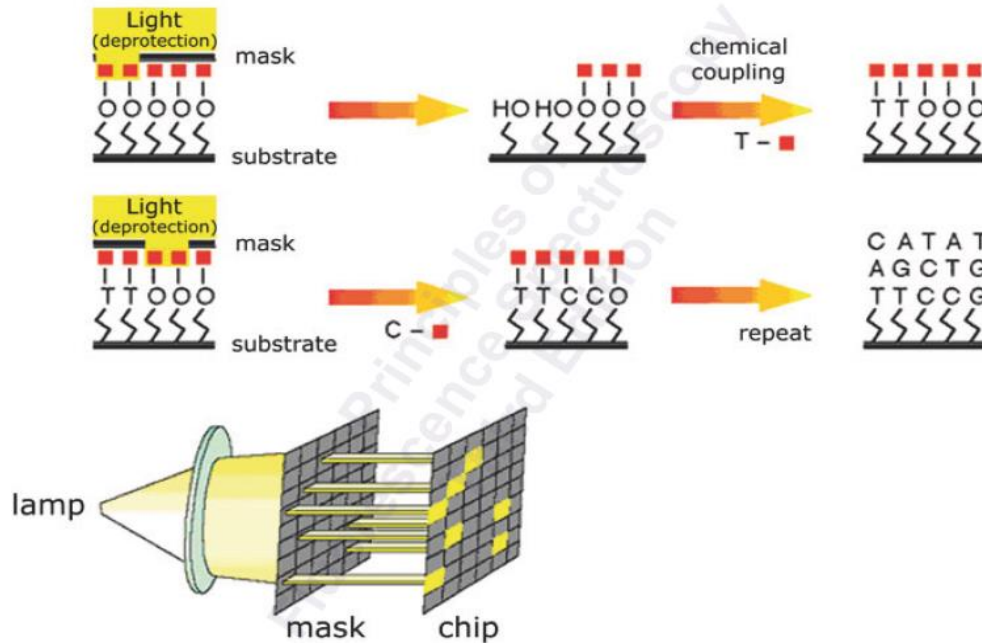


→ **In-situ** solid phase **synthesis** (directly on the chip) of DNA by photolithography

**Oligonucleotide Array**

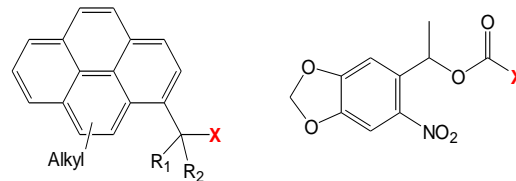
→ Affymetrix  
D.J. Lockhart et al.  
Nat. Biotechn. **1996**, 14, 1675

# Photolithographic in-situ Synthesis of Oligonucleotide Arrays



*Affymetrix*

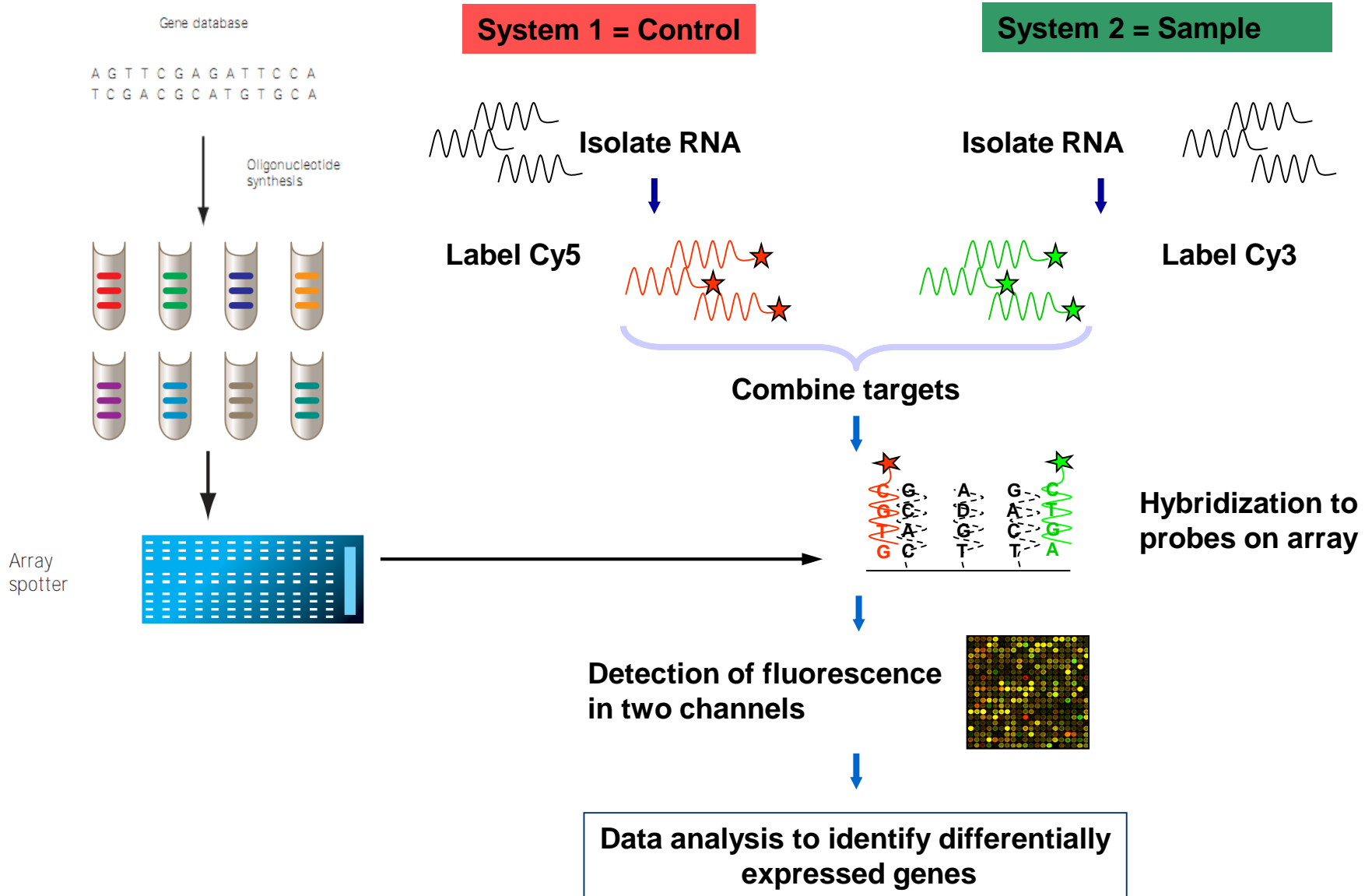
- light sensitive protecting groups, light + mask technique
- in situ combinatoric solid phase synthesis



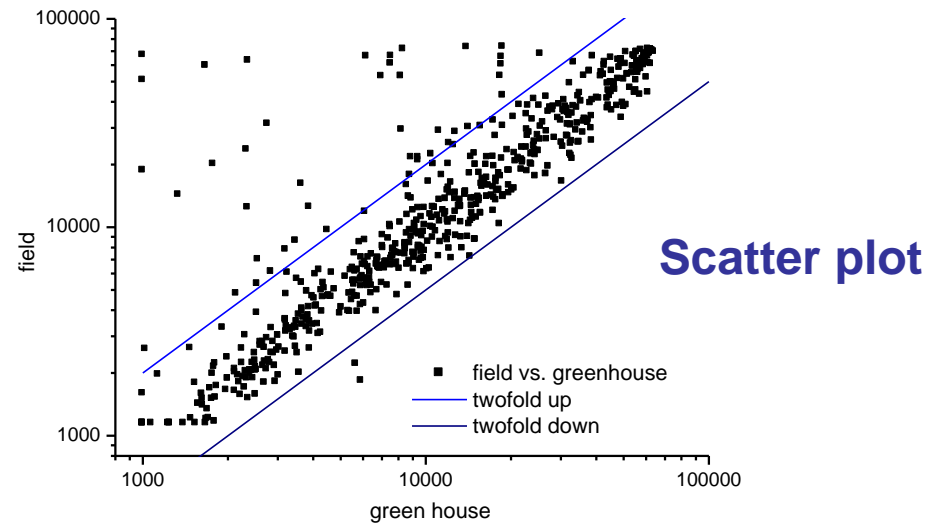
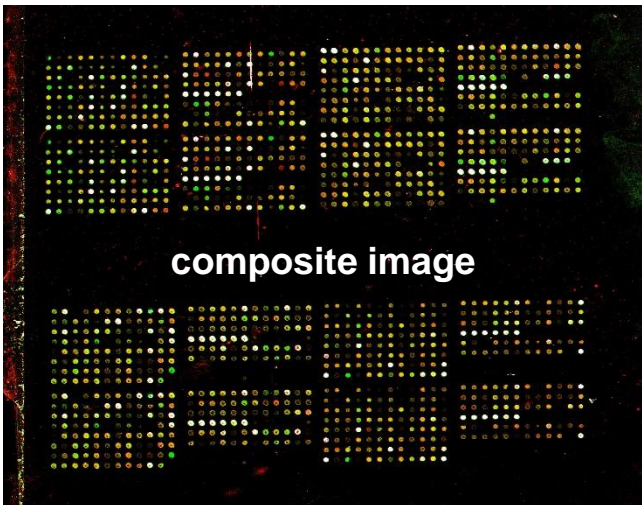
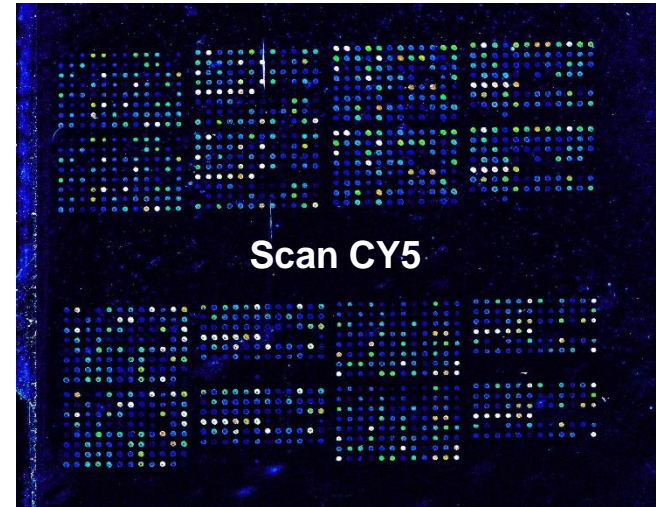
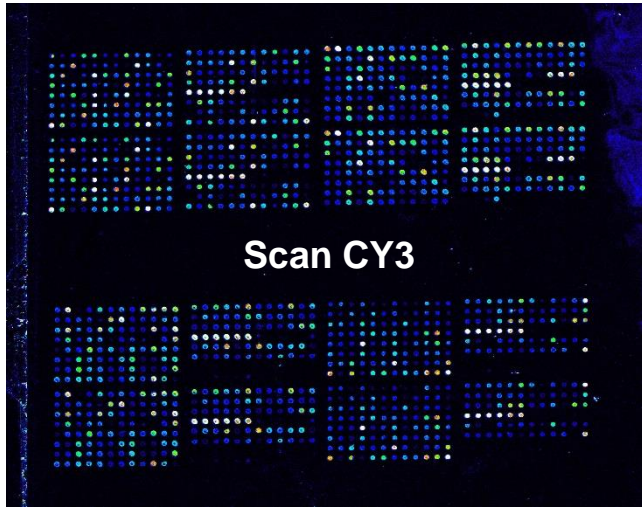
- incubation of activated areas with protected nucleotides
- 32 cycles (32 masks) → 65000 oligonucleotides (8 bases)



# Sample Preparation - RNA

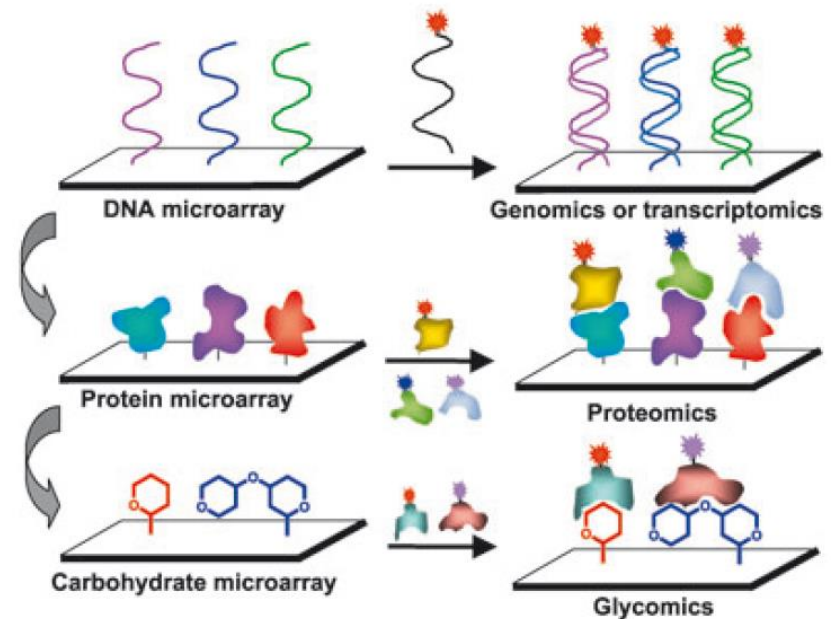


# Detection → Two Color Experiment

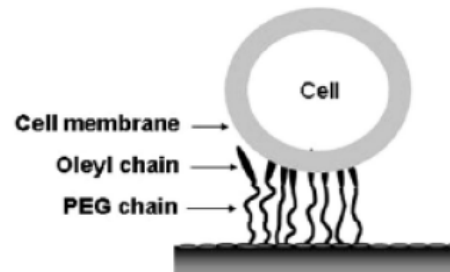


# DNA, Protein, Peptide, Carbohydrate, Cell, Tissue, ... Microarrays

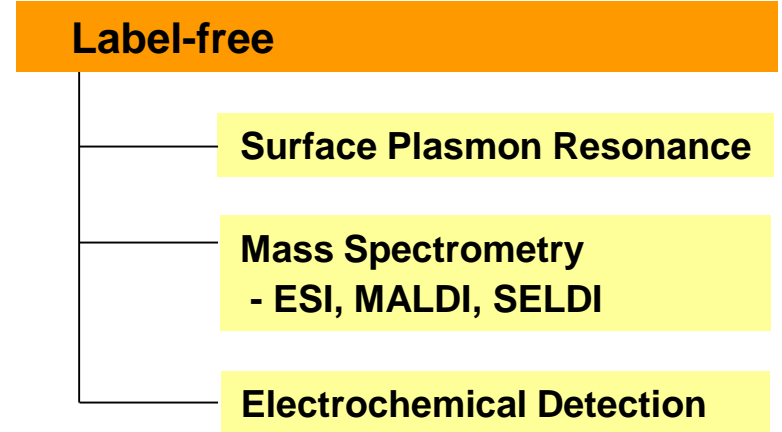
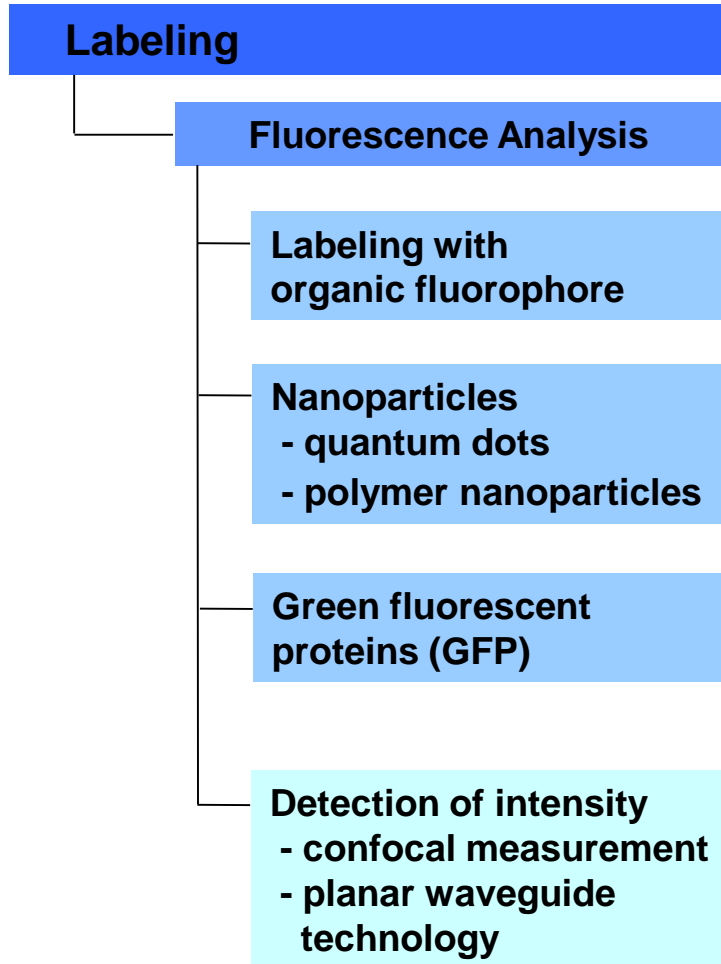
- DNA- and oligonucleotide microarrays
  - **Probes:** ssDNA, oligonucleotides
  - **Targets:** ssDNA, ssRNA
  - Nucleic acid „chemistry“
- Protein and peptide microarrays
  - **Probes:** Any protein (e.g. antigen)
  - **Targets:** Any other protein (e.g. antibody)
  - Amino acid „chemistry“



- Others
  - **Cell / tissue microarrays**
  - Transfection microarrays
  - ....

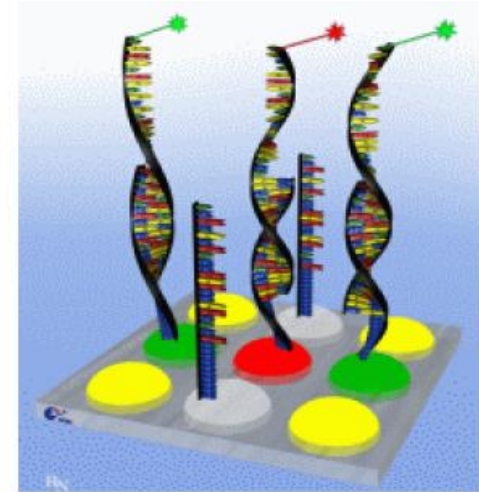
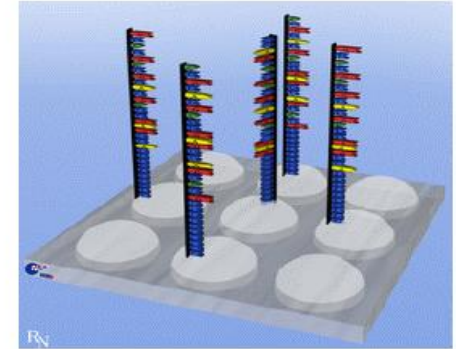


# Detection Principles for Microarrays

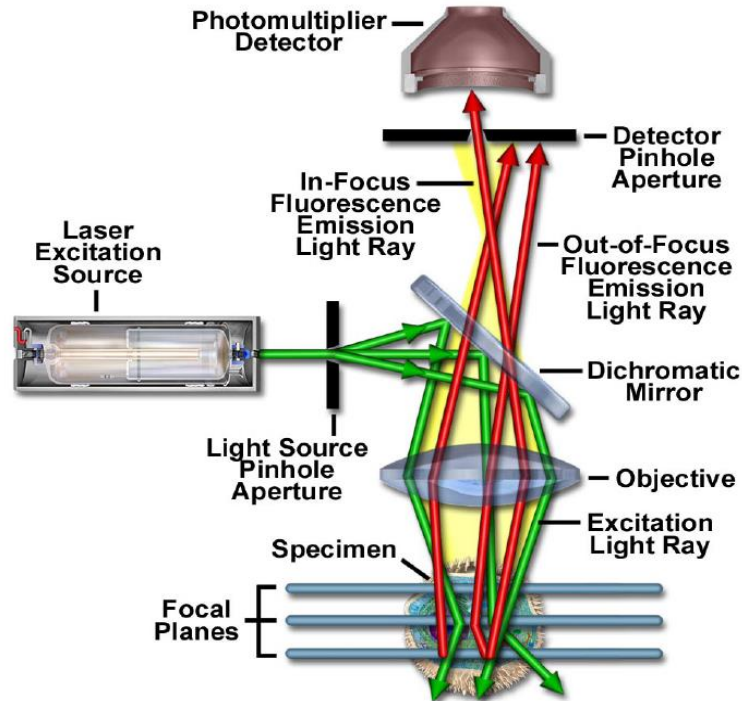


# Detection Principles for Microarrays

- Labeling of probe or target with fluorescent dye
  - Large amount of possible techniques known
  - Multiplexing possible (different dyes / colors)
  - Signal can be interpreted quantitatively (intensity  $\sim$  # hybridize molecules)
  - Fluorescent signal occurs only where hybridisation has taken place
- Detection of fluorescent signal with optical reader
- Practical issues
  - Unspecific binding
  - Photobleaching
  - Quenching



# Detection of Fluorescence using Principle of Confocal Microscopy



[www.zeiss.com](http://www.zeiss.com)

## Advantages

- Single pixel laser excitation
- Very good background suppression  
→ high S/N ratio

## Disadvantages

- Limited by choice of excitation wavelengths
- Focus depth of some 10  $\mu\text{m}$  requires substrates of high planarity and exact positioning
- High hardware costs

# Using of Laser Scanners



**Axon Genepix 4000B**

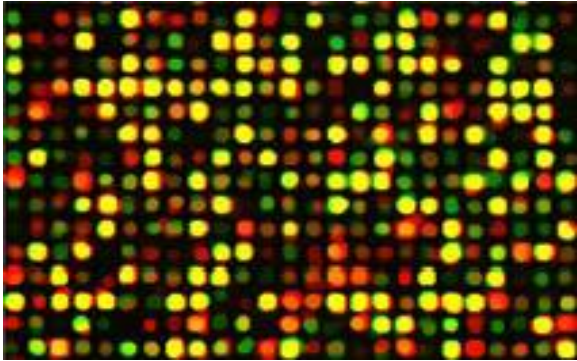
## Typical Features of a Laser Scanner:

- **Excitation** → Laser, e.g. solid state (488 nm), HeNe (543 nm, 594 nm, 633), Nd YAG (532 nm), 10-20 mW
- **Detection** → Photomultiplier tube or CCD camera
- **Sensitivity** → 0.02 – 1 fluorophor CY5 / mm<sup>2</sup>
- **Resolution** → 5 - 100 μm
- **Image format** → 16 or 24 bit tif

# Advantage of Confocal Measurement

## Planar supports

Confocal Scan



Non-Confocal Scan

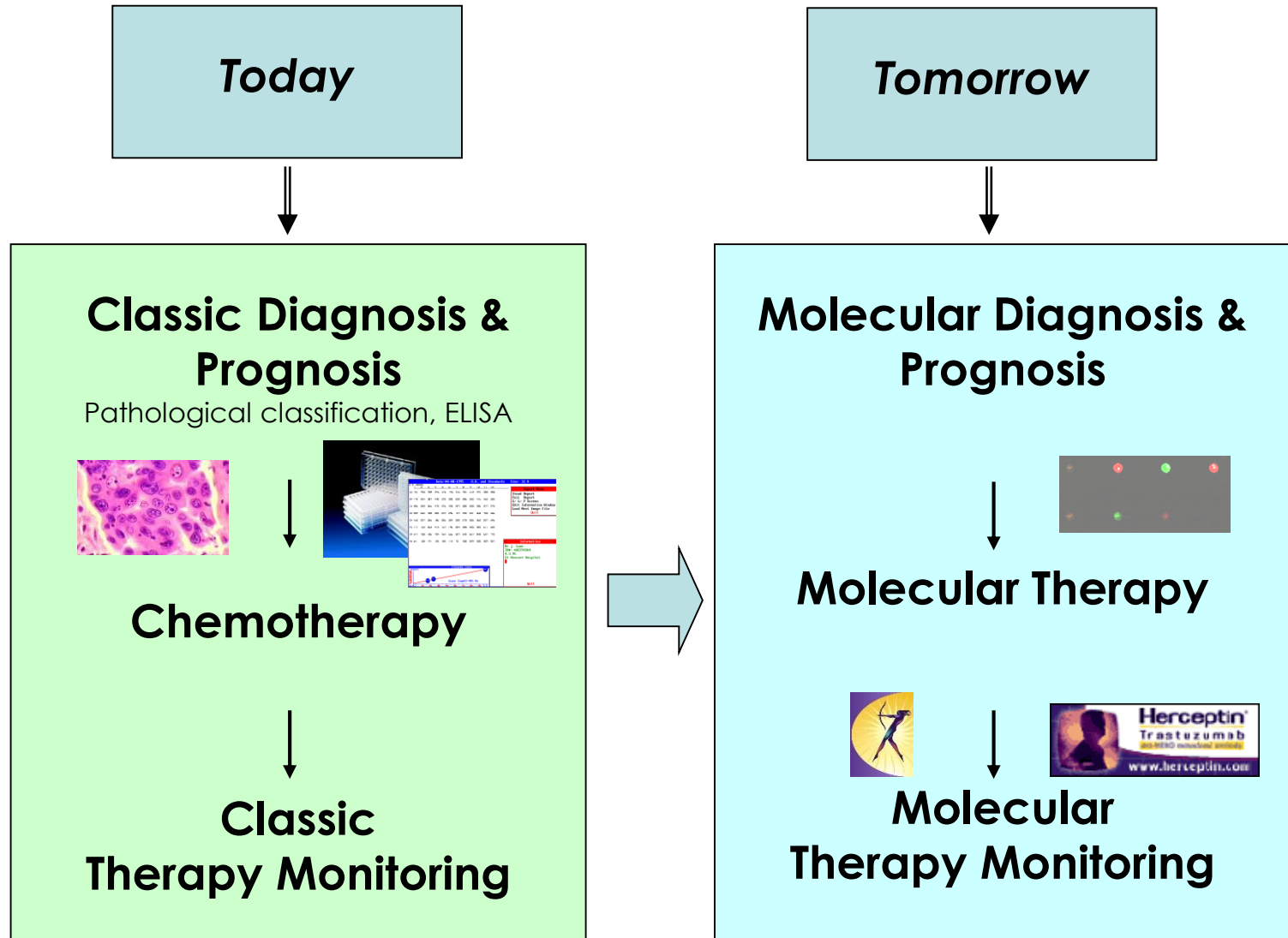


[www.tecan.com](http://www.tecan.com)

- ⇒ better signal to background ratio for planar surfaces and formats with 3D structure
- ⇒ especially for materials with intrinsic higher fluorescence background like polymers

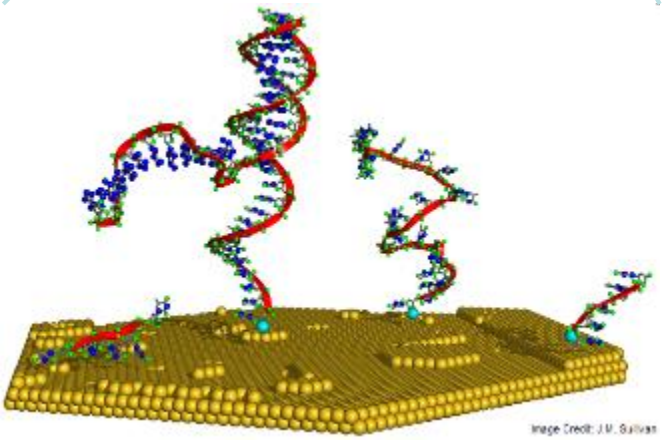
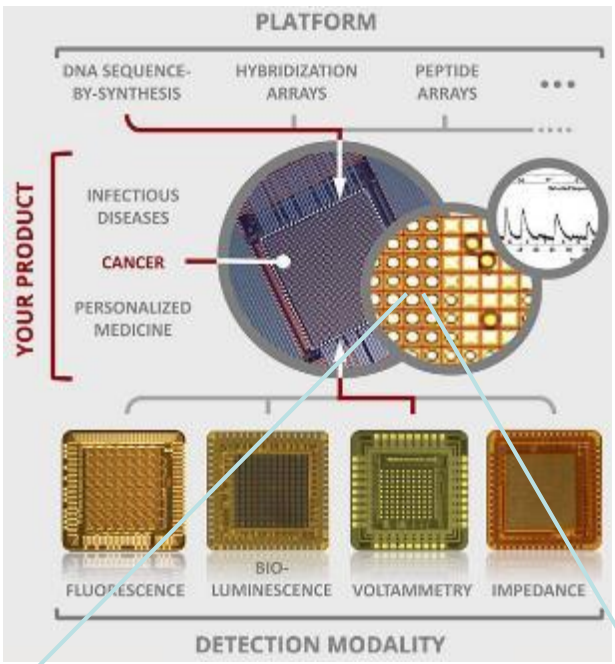
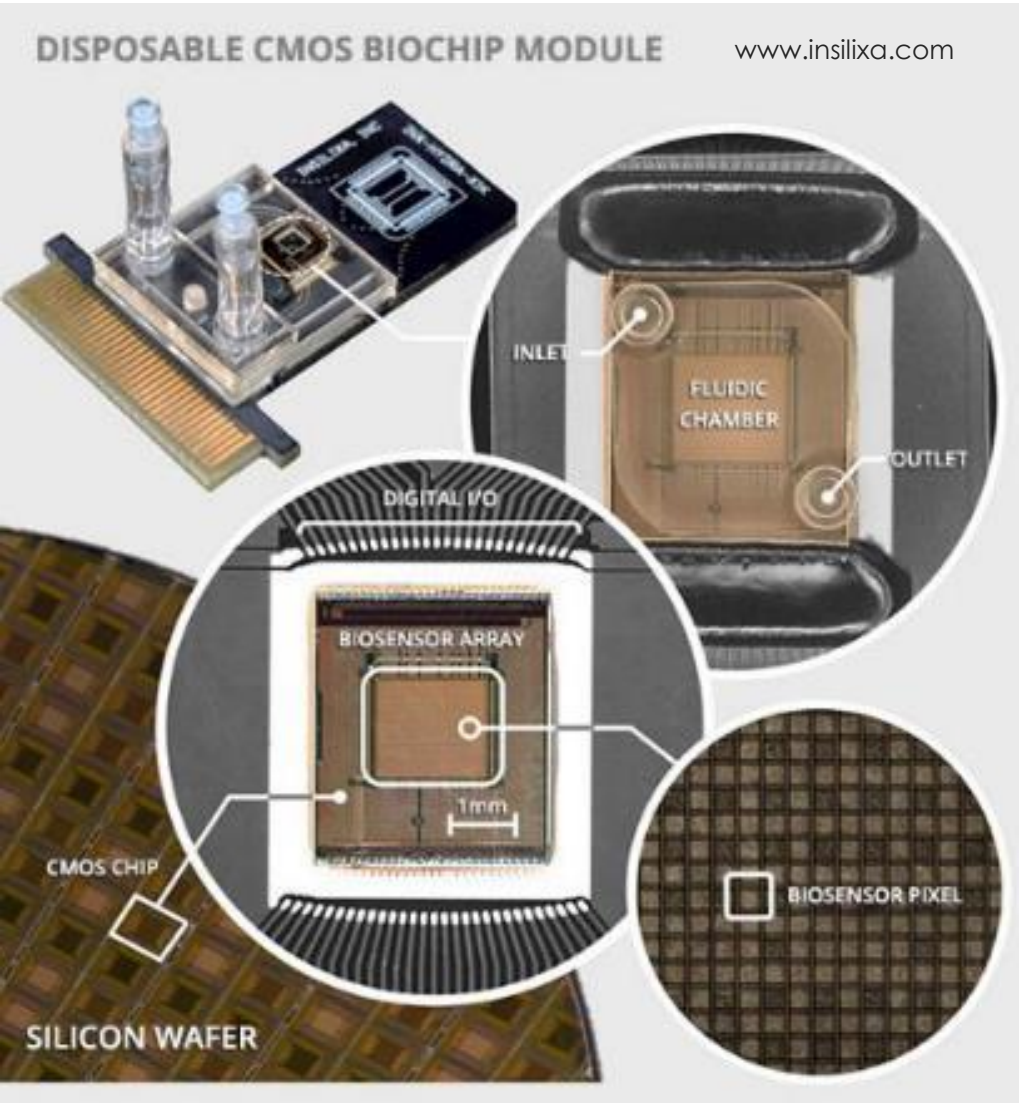


# Microarrays - Clinic



# **Biosensors**

# Biosensors Require Exact Loading of Sensor Fields



→ Control of behavior of biomolecules attached to surface on a nanometer scale

# Biosensors



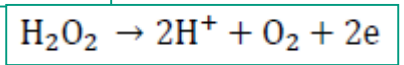
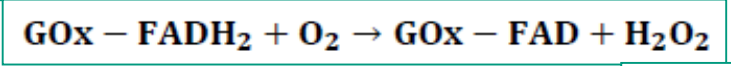
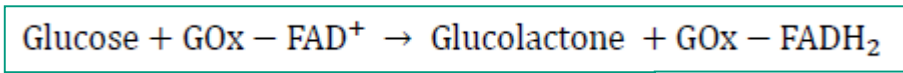
**Biosensor** → Combination of biological **recognition element** a **microfluidic chip** and a physical or chemical **transduces** to detect an **analyte**

Blood Glucose Measurement 85% of Market

# Glucose Sensing - Basic Principles

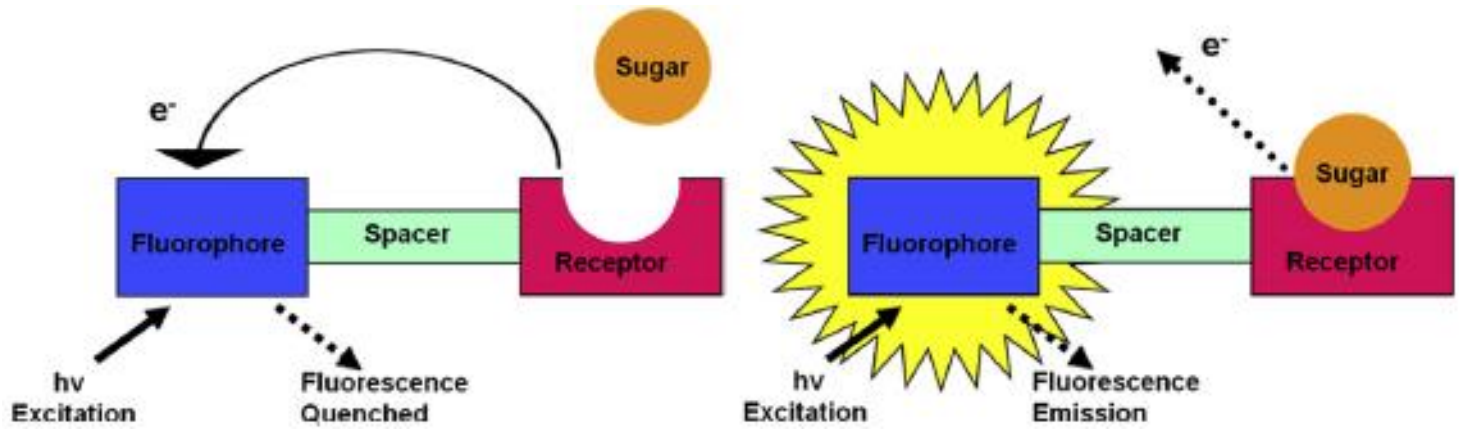
## Enzymes

- *Glucose Oxidase (GOx)*



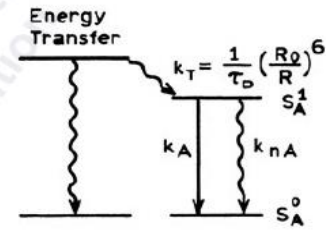
- Used in biosensors
- Easy to obtain, cheap
- High selectivity, withstands greater extremes of pH, T, ions

## Enzyme free



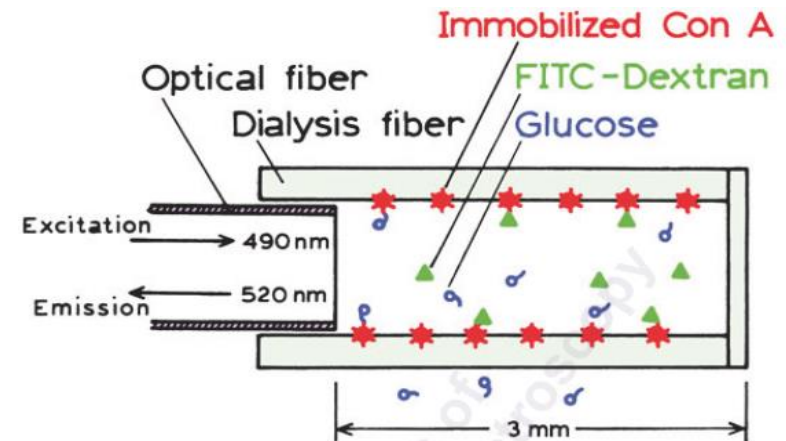
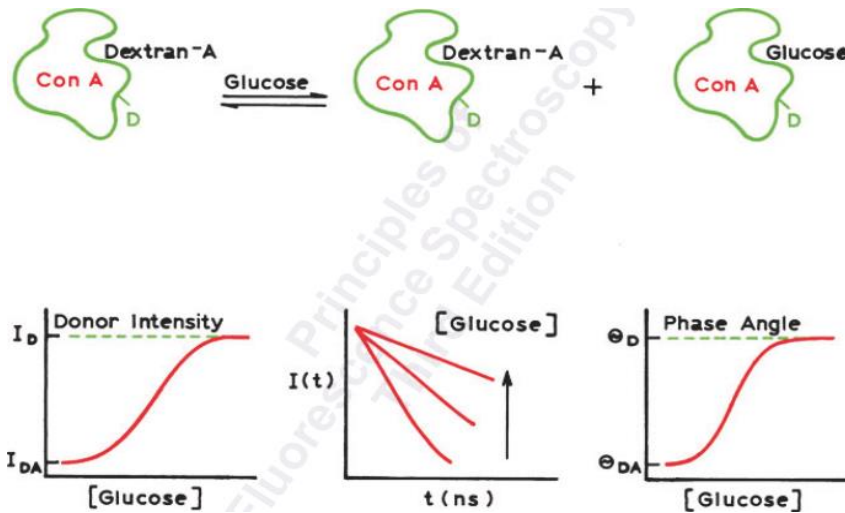
# Sensing basierend auf FRET

## → Blutzuckerbestimmung



### Glucose-Sensor

- Protein Concanavalin A (Con A) gelabelt mit Donor
- Glukose + konkurrierendes Polysaccharide (Dextran) gelabelt mit Akzeptor



- Bindung von Dextran-A an Con-A → Abnahme Donor FL-Intensität und Lebensdauer
- Bindung von Glucose → weniger Energietransfer, Zunahme FL-Signal vom Donor